

limiting factor for the crystal growth cannot now be determined.

Because aspartic acid is usually the most abundant residue in the SM of several kinds of molluscan shells (CRENSHAW, 1972; WEINER & HOOD, 1975; WEINER, 1979; NAKAHARA *et al.*, 1982), this acidic amino acid has been assumed to be one of the most important components involved in shell formation. WEINER (1979) and WEINER & TRAUB (1981) suggested that the SM may form a two-dimensional sheet with regularly spaced carboxyl groups of aspartic acids on the surface. This precise spatial arrangement of the carboxyl-groups may react with  $\text{Ca}^{2+}$  ions and promote the nucleation of crystals of  $\text{CaCO}_3$ . DEGENS (1979) presented a similar idea in relation to the role of the SM. As in the previous data, which were based on the measurements made on the unfractionated matrix, aspartate was usually the most abundant residue in the purified Ca-binding glycoproteins, except in the complex and crossed-lamellar layers. However, the amount of aspartate showed distinctive variations according to the ultrastructure of the shell. The most distinctive difference was between the nacreous and prismatic layers, and the homogeneous, composite-prismatic, complex, and crossed-lamellar layers. This result could imply different functions of the glycoproteins in the process of shell formation. In the foliated and chalky layers, aspartate was present in high amounts but serine was also highly concentrated. Serine is generally recognized to be present mostly as phosphoserine (BUTLER, 1987). WHEELER *et al.* (1988) demonstrated that such phosphorylation of the matrix may be significant for regulating the morphology of carbonate. The amino acid compositions of the organic matrix may also depend on various environmental factors to which mollusks are subjected. From the results of a comparative analysis on the unfractionated matrix, DEGENS *et al.* (1967) showed that amino acid compositions of the matrix are correlated with environmental factors. DUSSART (1984) also reported that the amino acid compositions of the shells of 13 species of freshwater Bivalvia reflected phylogenetic affinity but that environmental factors were probably important. Samata (unpublished data) has also pointed out the slight difference in the amino acid compositions of the Ca-binding glycoproteins in the nacreous and prismatic layers between the marine and freshwater species of Bivalvia. The difference was most remarkable with respect to the levels of aspartate.

The compositions of the Ca-binding glycoproteins in the nacreous and prismatic layers of *Nautilus pompilius* were slightly different from those in the same layers of the species of Gastropoda and Bivalvia, and also from those in the other shell layers. *Nautilus* is distributed in fairly deep waters in tropical seas, whereas the other species examined are restricted to shallow waters. The unique habitat of *Nautilus* makes it difficult to determine whether the difference in amino acid composition can be accounted for by environmental or phylogenetic factors.

The amino acid composition of the Ca-binding glycoprotein in the composite-prismatic layer was clearly different from those of the nacreous, prismatic, and foliated layers. The composite-prismatic layer was first defined by BØGGILD (1930) and has been considered to be a subdivision of the prismatic layer by some later investigators (CARTER, 1980; UOZUMI & SUZUKI, 1981). KOBAYASHI (1968) noted that the ISM in this layer resembled that in the complex and crossed-lamellar layers both morphologically and histochemically, but differed from that in the nacreous and prismatic layers. Moreover, because the mollusks that contain this layer are taxonomically close to those that contain the homogenous, complex, and crossed-lamellar layers, the composite-prismatic layer may be closely related to these three shell layers.

#### ACKNOWLEDGMENTS

I am grateful to Prof. M. Omori, Faculty of General Education, Azabu University, for his most helpful comments during the preparation of the manuscript and the provision of the specimens of *Peronidia* and *Glycymeris*. I thank also Prof. N. Watabe, Department of Biology, University of South Carolina, for reviewing the manuscript and for his invaluable suggestions. This research was partly supported by a Grant-in-Aid for Scientific Research from the Ministry of Education and Culture of Japan.

#### LITERATURE CITED

- AKIYAMA, M. 1966. Conchiolin-constituent amino acids and shell structures of bivalved shells. *Proc. Japan Acad.* 42(7): 800-805.
- ANDERSON, B. L., R. W. BERRY & A. TELSER. 1983. A sodium dodecyl sulfate-polyacrylamide gel electrophoresis system that separates peptides and proteins in the molecular weight range of 2,500 to 90,000. *Anal. Biochem.* 132:365-375.
- BØGGILD, O. B. 1930. The shell structure of the mollusks. *Kgl. Danske Videnskab Selsk. Skr. Nature* 9(2):233-326.
- BUTLER, W. T. 1987. Mineralized tissues: an outview. *Met. Enzymol.* 145:255-261.
- CARTER, J. G. 1980. Guide to bivalve shell microstructures. Pp. 645-673. *In*: D. C. Rhoades & R. A. Lutz (eds.), *Skeletal growth of aquatic organisms*. Plenum Press: New York.
- CARTER, J. G. & G. R. CLARK, II. 1985. Classification and phylogenetic significance of molluscan shell microstructure. Pp. 50-71j. *In*: T. W. Broadhead (ed.), *Mollusks, Notes for a short course*. Dept. Geol. Sci., Univ. Tennessee: Knoxville.
- CRENSHAW, M. A. 1972. The soluble matrix from *Mercenaria mercenaria* shell. *Biomineralization* 6:6-11.
- DEGENS, E. T. 1979. Molecular mechanisms of carbonate, phosphate and silica deposition in the living cell. *Top. Curr. Chem.* 64:1-112.
- DEGENS, E. T., D. W. SPENCER & R. H. PARKER. 1967. Paleobiochemistry of molluscan shell proteins. *Comp. Biochem. Physiol.* 20:533-579.
- DUSSART, G. B. J. 1984. The amino acid composition of fresh water mollusc shells in relation to phylogeny and environment. *Jour. Molluscan Stud.* 49:213-223.
- FREMY, M. 1855. Recherches chimique sur les os. *Ann. Chem. Phys.* 43:96.

- GRÉGOIRE, C. 1972. Structures of the molluscan shell. Pp. 45-102. *In*: M. Florkin & B. T. Scheer (eds.), Chemical zoology, Vol. II, Mollusca. Academic Press: London.
- KASAI, H. & N. OHTA. 1981. Relationship between organic matrices and shell structures in recent bivalves. Pp. 101-106. *In*: T. Habe & M. Omori (eds.), Studies of molluscan paleobiology. Professor Masae Omori Memorial Volume Publication Committee, Niigata Univ. Press: Niigata.
- KOBAYASHI, I. 1968. The relation between the morphological structure types of shell tissues and the nature of the matrices in the bivalve molluscs. *Venus* 27-3:111-123.
- KOBAYASHI, I. 1971. Internal shell microstructure of recent bivalvian molluscs. *Sci. Rept. Niigata Univ.* E(2):27-50.
- KRAMPITZ, G. & W. WITT. 1979. Biochemical aspects of biomineralization. *Top. Curr. Chem.* 78:57-144.
- KRAMPITZ, G., J. ENGELES & C. CAZAUX. 1976. Biochemical studies on water soluble proteins and related components of gastropod shells. Pp. 155-173. *In*: N. Watabe & K. M. Wilbur (eds.), The mechanism of mineralization in the invertebrates and plants. Univ. South Carolina Press: Columbia.
- MACCLINTOCK, C. 1967. Shell structure of patelloid and bellerophonid gastropods (Mollusca). *Peabody Mus. Natur. Hist., Yale Univ. Bull.* 22:1-140.
- MEENAKSHI, V. R., P. E. HARE & K. M. WILBUR. 1971. Amino acids of organic matrix of neogastropod shell. *Comp. Biochem. Physiol.* 40B:1034-1043.
- NAKAHARA, H., G. BEVELANDER & M. KAKEI. 1982. Electron microscopic and amino acid studies of the outer and inner shell layers of *Haliotis rufescens*. *Venus* 41:33-46.
- O'FARRELL, P. H. 1975. High resolution two-dimensional electrophoresis of proteins. *Jour. Biol. Chem.* 250(10):4007-4021.
- SAMATA, T. 1988a. Studies on the organic matrix in molluscan shells. I. Amino acid composition of the organic matrix in the nacreous and prismatic layers. *Venus* 47(2):127-140.
- SAMATA, T. 1988b. Biochemical studies on the organic matrix in hard tissues. 2. The structure of the insoluble organic matrix in the foliated, homogeneous, composite-prismatic, complex and crossed-lamellar layers of molluscan shells. *Jour. Fac. General Education Azabu Univ.* 21:71-82.
- SAMATA, T. & G. KRAMPITZ. 1981. Ca-binding polypeptides in oyster shells. *Malacologia* 22:225-233.
- SAMATA, T. & M. MATSUDA. 1986. Contaminating peptides widely present in ion-exchanged water, reagents, experimental implements and natural sample. *Comp. Biochem. Physiol.* 84B:200-212.
- SCHIELDS, R. & W. BURNET. 1960. Determination of protein-bound carbohydrate in serum by a modified anthron method. *Anal. Chem.* 32:885-886.
- TAUSSKY, H. H. & E. SCHORR. 1967. A microcolorimetric method for the determination of inorganic phosphorus of invertebrate mineralized tissues. *Jour. Ultrastruc. Res.* 18: 519-550.
- TAYLOR, J. D., J. M. KENNEDY & A. HALL. 1969. The shell structure and mineralogy of the Bivalvia—Introduction, Nuculacea-Trigonacea. *Bull. Br. Mus. Natur. Hist. (Suppl.)* 3:1-125.
- UOZUMI, S. & S. SUZUKI. 1981. The evolution of shell structure in the Bivalvia. Pp. 63-77. *In*: T. Habe & M. Omori (eds.), Studies of molluscan paleobiology. Professor Masae Omori Memorial Volume Publication Committee, Niigata Univ. Press: Niigata.
- WADA, K. 1964. Studies on the mineralization of calcified tissue in molluscs. VII. Histological and histochemical studies of organic matrices in shells. *Bull. Natl. Pearl Res. Lab.* 9: 1078-1086.
- WADA, K. 1980. Initiation of mineralization in bivalve molluscs. Pp. 79-92. *In*: M. Omori & N. Watabe (eds.), The mechanism of biomineralization in animals and plants. Tokai Univ. Press: Tokyo.
- WEINER, S. 1979. Aspartic acid rich proteins: major component of the soluble organic matrix of mollusc shells. *Calcif. Tissue Int.* 29:163-167.
- WEINER, S. 1983. Mollusk shell formation: isolation of two organic matrix proteins associated with calcite deposition in the bivalve *Mytilus californianus*. *Biochemistry* 22:4139-4144.
- WEINER, S. & L. HOOD. 1975. Soluble protein of the organic matrix of mollusc shells: a potential template for shell formation. *Science* 190:887-898.
- WEINER, S. & W. TRAUB. 1981. Organic-matrix-mineral relationship in mollusk shell nacreous layers. Pp. 467-482. *In*: M. Balaban, J. L. Sussman, W. Traub & A. Yonath (eds.), Structural aspects of recognition and assembly in biological macromolecules. Balaban I SS, Rehovot: Philadelphia.
- WEINER, S., H. A. LOWENSTAM & L. HOOD. 1977. Discrete molecular weight components of the organic matrices of molluscs. *Jour. Exp. Mar. Ecol.* 30:45-51.
- WHEELER, A. P., K. W. RUSENKO & C. S. SIKES. 1988. Organic matrix from carbonate biomineral as a regulator of mineralization. Pp. 9-13. *In*: C. S. Sikes & A. P. Wheeler (eds.), Chemical aspects of regulation of mineralization. Univ. South Alabama Publication Series: Alabama.

A New Species of the Genus *Cyerce* Bergh, 1871,  
from the Cape Verde Islands  
(Opisthobranchia: Ascoglossa)

by

JESÚS ORTEA

Departamento de Biología, Facultad de Ciencias,  
Universidad de Oviedo, Oviedo, Spain

AND

JOSÉ TEMPLADO

Museo Nacional de Ciencias Naturales (C.S.I.C.),  
José Gutiérrez Abascal 2, 28006 Madrid, Spain

*Abstract.* *Cyerce verdensis* Ortea & Templado, sp. nov., is described from the Cape Verde Islands (eastern Atlantic). The presence of papillae on the rhinophores, pericardium, and cerata is the main differential feature of this new species. The Atlantic species of the genus *Cyerce* are listed and discussed.

INTRODUCTION

In a recent paper (ORTEA & TEMPLADO, 1988) a new species from Cuba, *Cyerce habanensis*, was described, and the Atlantic species of this genus were discussed. At the same time, THOMPSON (1988) described another new species of this genus from the Saronic Gulf, *C. graeca*, in one of his works on eastern Mediterranean opisthobranchs. Following his advice on the necessity of further studies on this genus, we describe here another new species of *Cyerce* from the tropical eastern Atlantic, collected during the "Primera Expedición Científica Ibérica al Archipiélago de Cabo Verde" (August of 1985).

Family CALIPHYLLIDAE Thiele, 1931

Genus *Cyerce* Bergh, 1871  
= *Lobifera* Pease, 1866  
= *Lobiancoia* Trinchese, 1881

*Cyerce verdensis* Ortea & Templado, sp. nov.

(Figures 1-3)

**Material:** One specimen (Figures 1-3), 16 mm in length (18 August 1985), and four others 13, 12, 10, and 5 mm

in length (20 August 1985), all collected between 0 and 2 m deep on *Halimeda* sp., in Salamanca Bay (16°54'N, 24°57'W), San Vicente Island, Cape Verde Archipelago.

The largest specimen has been chosen as holotype, and deposited in the Museo Insular de Ciencias Naturales, Tenerife, Canary Islands (catalogue number MO/0092). The four paratypes are in the malacological collection of the Museo Nacional de Ciencias Naturales de Madrid, Spain (catalogue number 15.05/1033).

**Etymology:** Named after the Cape Verde Islands.

**Description:** The general body color of the animal is pale ochre. The dark brown digestive gland, which can be discerned clearly through the skin, is divided into two main branches that almost reach the posterior part of the dorsum. Each branch ramifies towards the cerata, without extending into them (Figure 1).

The foot is light in color, almost semi-translucent. Its frontal margin is rounded and a transverse mesopodial groove is present in the anterior one-third (Figure 2A).

The brownish pericardium has granulose white papillae. Such small papillae are present all over the dorsum. The bulky anal papilla is situated just before the pericardium, somewhat to the right of the median plane.

The rhinophores are split and inrolled. They have a disperse, brown and white pigmentation. Granulose papillae of a pale creme color are present on their distal half. The oral tentacles, also inrolled, are translucent with some white dots.

The leaf-shaped cerata are translucent with their distal margins angulose owing to some small white granules. Papillae, white granules, and superficial red-brown specks are present on both sides of the cerata (Figure 2B, C).

The radula of the 16-mm long specimen has 10 teeth in the ascending series and 13 in the descending one. The ascus contains more than 100 cluttered teeth. The functional teeth reach up to 80  $\mu\text{m}$  in length at the beginning of the descending series. They are elongate and exhibit 12 denticles on both cutting edges. The protuded median zone of each tooth has two blunt protuberances (Figure 3B).

The penis is armored with a small nail-like spine that measures 10  $\mu\text{m}$  in length (Figure 3A).

**Discussion:** In a former paper (ORTEA & TEMPLADO, 1988) we have already discussed the Atlantic species of this genus. *Cyerce verdensis* clearly differs from all these by the presence of granulose papillae all over the dorsum, rhinophores, and cerata. Such papillae are lacking in the other species. The disposition of the papillae in the cerata resembles *Polybranchia viridis* (Deshayes, 1857), previously collected in the Canary Islands (ORTEA, 1981). This latter species reaches 55–70 mm in length, and its juveniles are easily confused with *C. verdensis* at first sight. However, *P. viridis* lacks a transverse mesopodial groove and the diverticula of the digestive gland extend into the cerata.

The radular teeth of *Cyerce verdensis* are shorter and wider in proportion compared to those of *C. cristallina* and *C. antillensis*, and similar to those of *C. graeca* and *C. habanensis*.

*Cyerce edmundsi* Thompson, 1977, from Jamaica, also exhibits some pearl-like white glands in the distal margin of the ceras. However, the lack of a transverse mesopodial groove and the diverticula of the digestive gland in the cerata suggests that it belongs to another genus. MARCUS (1982) and JENSEN & CLARK (1983) commented that *C. edmundsi* could be a junior synonym of *Mourgona germainae* Marcus & Marcus, 1970.

The fact that all the specimens of *Cyerce verdensis* were found on the chlorophyte *Halimeda* sp. suggests that this seaweed constitutes its food. We previously reported another species of this genus, *Halimeda opuntia*, as the probable food of *C. habanensis* (ORTEA & TEMPLADO, 1988).

Including the present paper, the Atlantic (including Mediterranean Sea) species of the genus *Cyerce* are the following:

- C. cristallina* (Trinchese, 1881). Mediterranean and Caribbean Sea.
- C. antillensis* Engel, 1927. Caribbean region.
- C. graeca* Thompson, 1988. Eastern Mediterranean (known from the Saronic Gulf only).

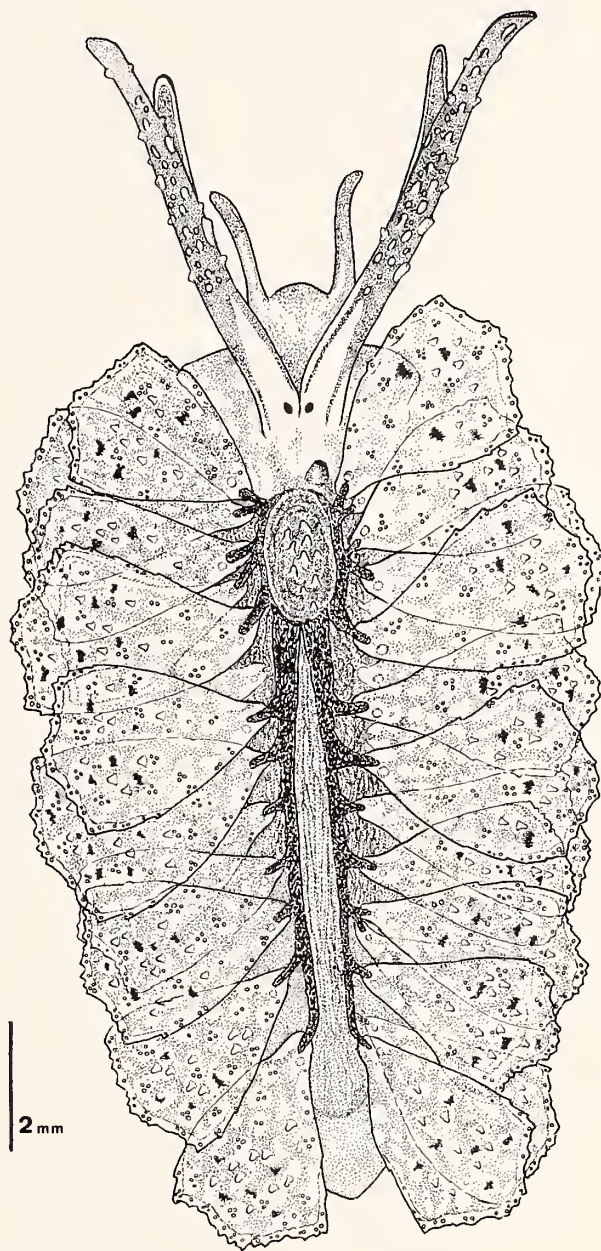


Figure 1

*Cyerce verdensis* Ortea & Templado, sp. nov., dorsal view of holotype.

- C. habanensis* Ortea & Templado, 1988. Caribbean region (known from northern Cuba only, but the species cited by MARCUS & HUGHES (1974) as *C. antillensis* in Barbados might be this species).
- C. verdensis* sp. nov. Cape Verde Islands (eastern Atlantic).

This last species is the first one of the genus which has been recorded from the eastern Atlantic Basin outside of the Mediterranean Sea.